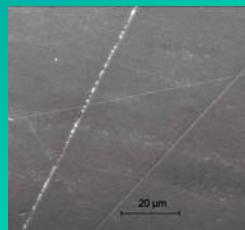


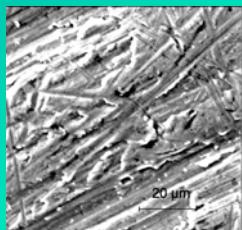
Methods for the physical and chemical characterisation of surfaces of titanium implants

Surface modifications of titanium implants

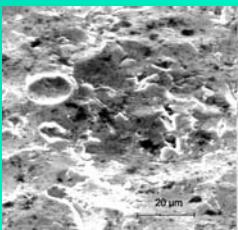
Polished
(P)



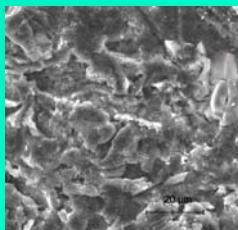
Machined
(M)



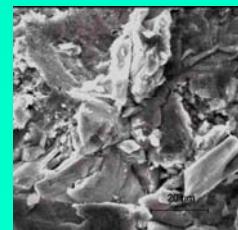
Glass-particle
blasted
(G)



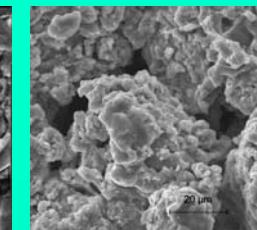
Corundum-
blasted 2,5 bar
(C2.5)



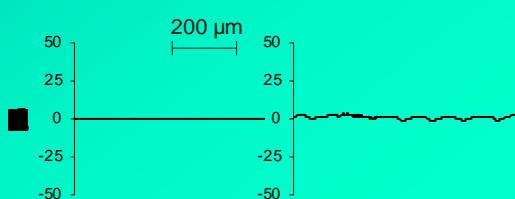
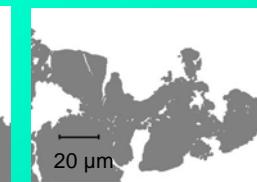
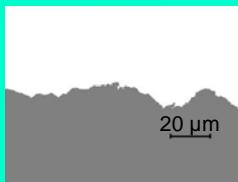
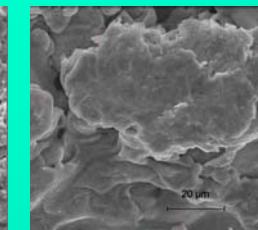
Corundum-
blasted 6 bar
(C6)



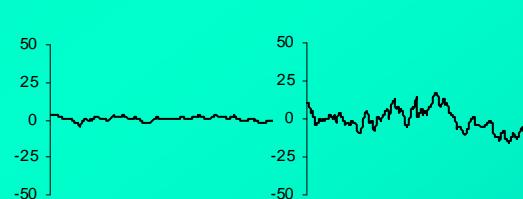
Vacuum-
plasma sprayed
rough (VR)



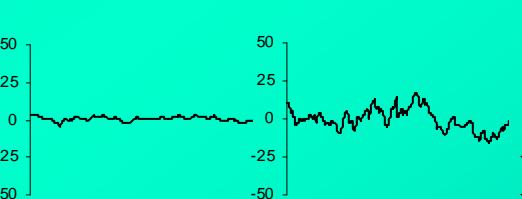
Vacuum-
plasma sprayed
fine (VF)



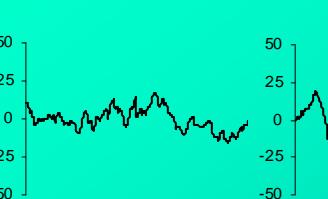
$R_a=0.07\mu\text{m}$



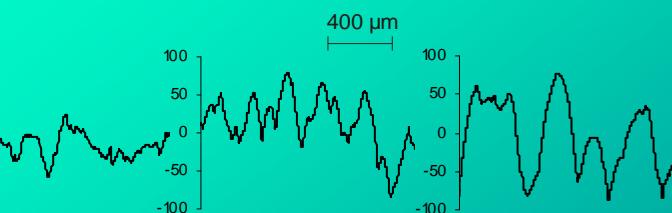
$R_a=0.53\mu\text{m}$



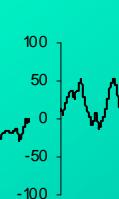
$R_a=1.22\mu\text{m}$



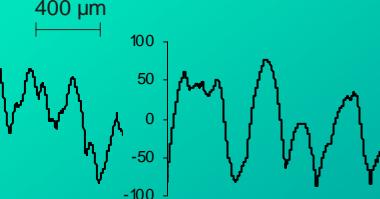
$R_a=4.12\mu\text{m}$



$R_a=6.07\mu\text{m}$



$R_a=20.99\mu\text{m}$



$R_a=48.59\mu\text{m}$

Commercial distributed implants do not have an uniform surface:
--> every part of the implant needs an **optimized** surface

Complete and exact
description of titanium surfaces with different roughness

Searching for relevant parameters for a correlation
physico-chemical characteristics --> cell behaviour
--> biocompatibility

(1) Estimation of the “true” surface area

quality control, input parameter for further working steps
in modifying the implant surface:

- electrochemical deposition of hydroxyapatite
- embedding of antibiotic substances

3 electrochemical methods:

- Electrochemical Impedance Spectroscopy (EIS)
- Linear Sweep Voltammetry (LSV)
- Chronoamperometry (CA)

(2) Characterisation of the surface roughness

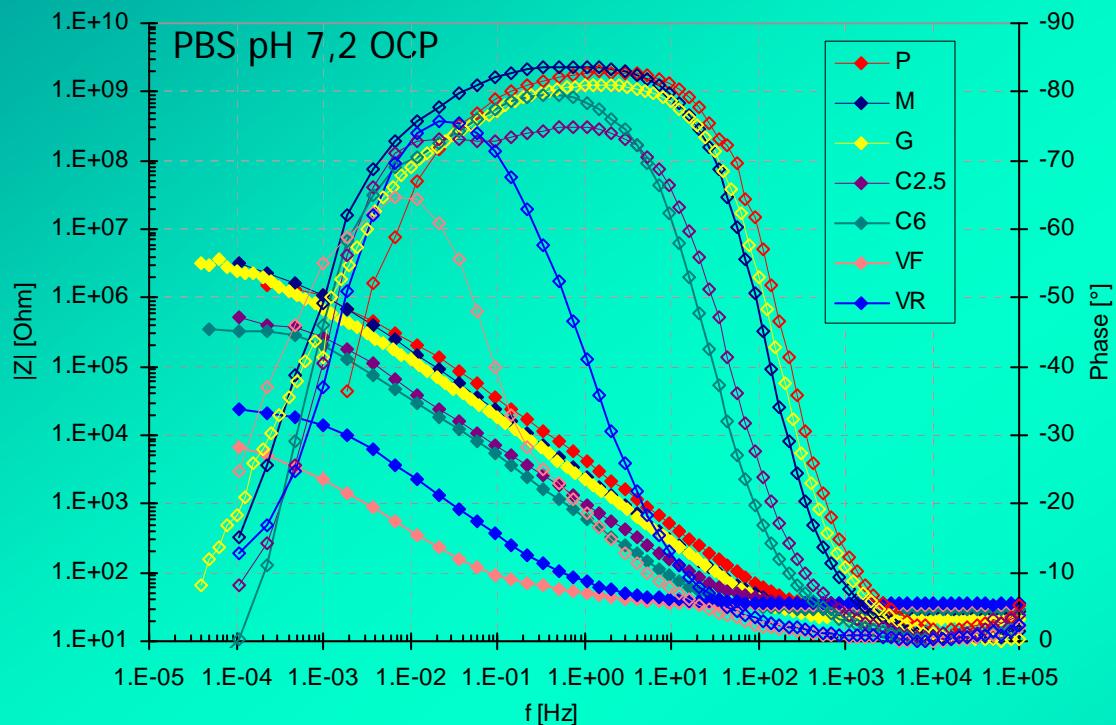
with the parameter **fractal dimension D_F**

-2 electrochemical methods: EIS and LSV
comparison with results of Digital Image Processing (DIP)

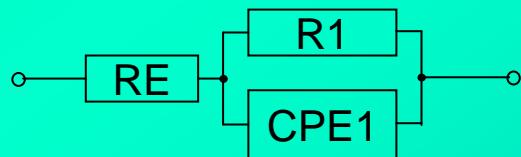
(3) Cell biological tests

Electrochemical Impedance Spectroscopy (EIS)

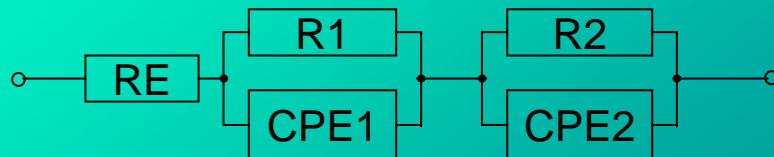
Bode plot for all investigated titanium surfaces



P, M, G, C2.5, C6



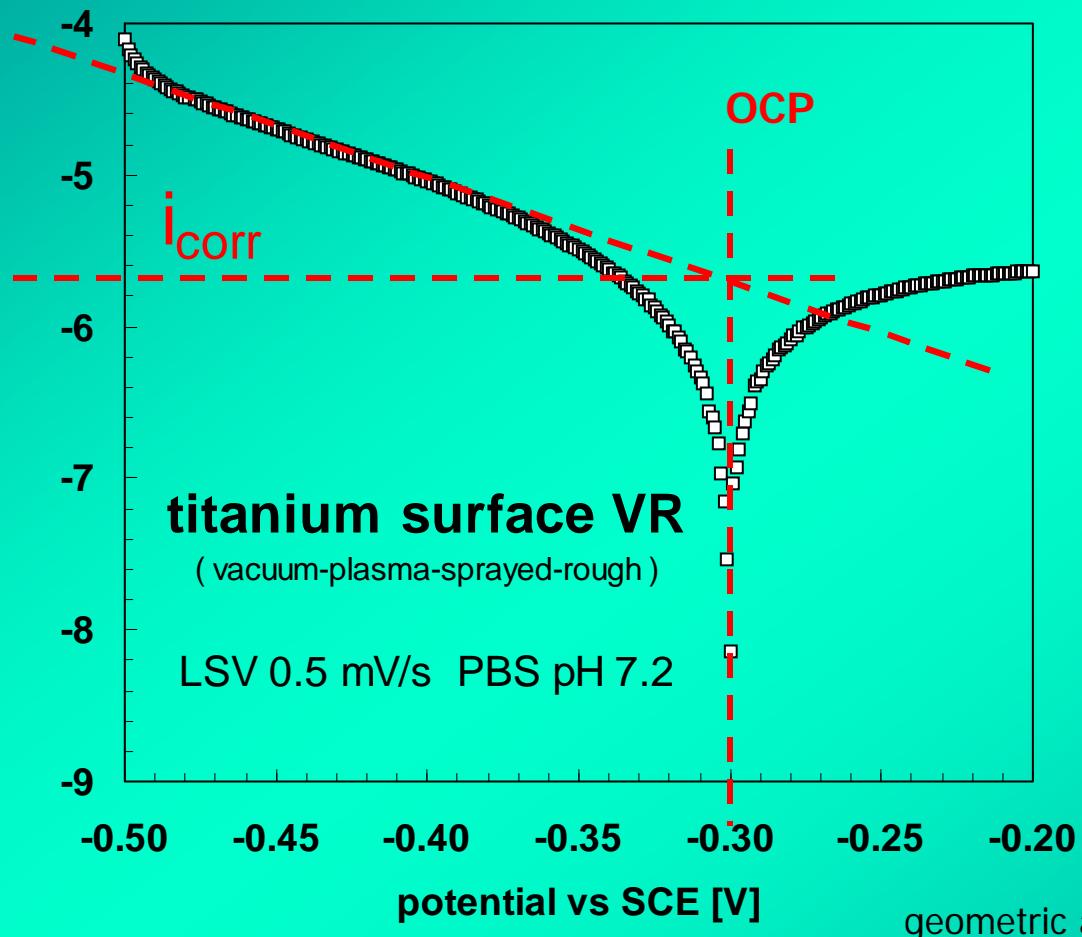
VF, VR



Linear Sweep Voltammetry (LSV)

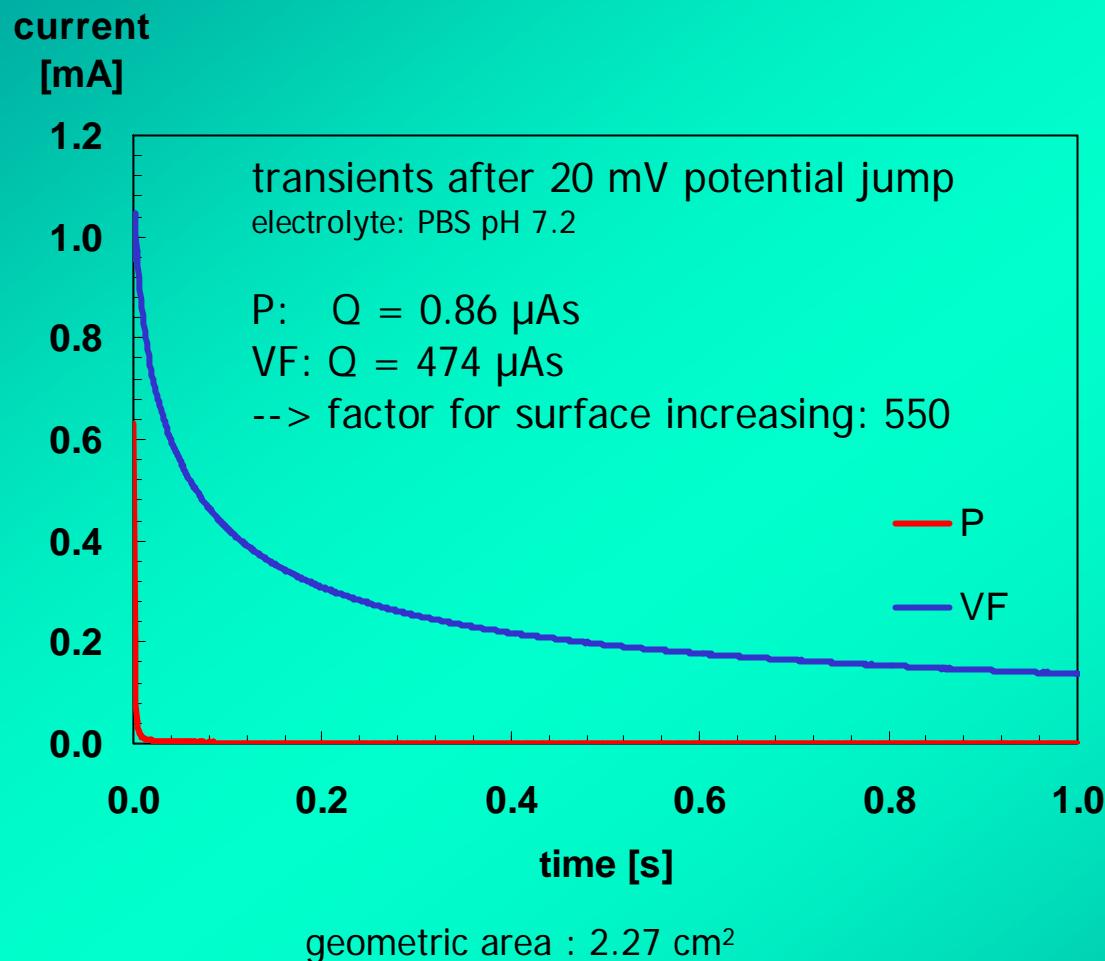
log current [A]

Tafel analysis



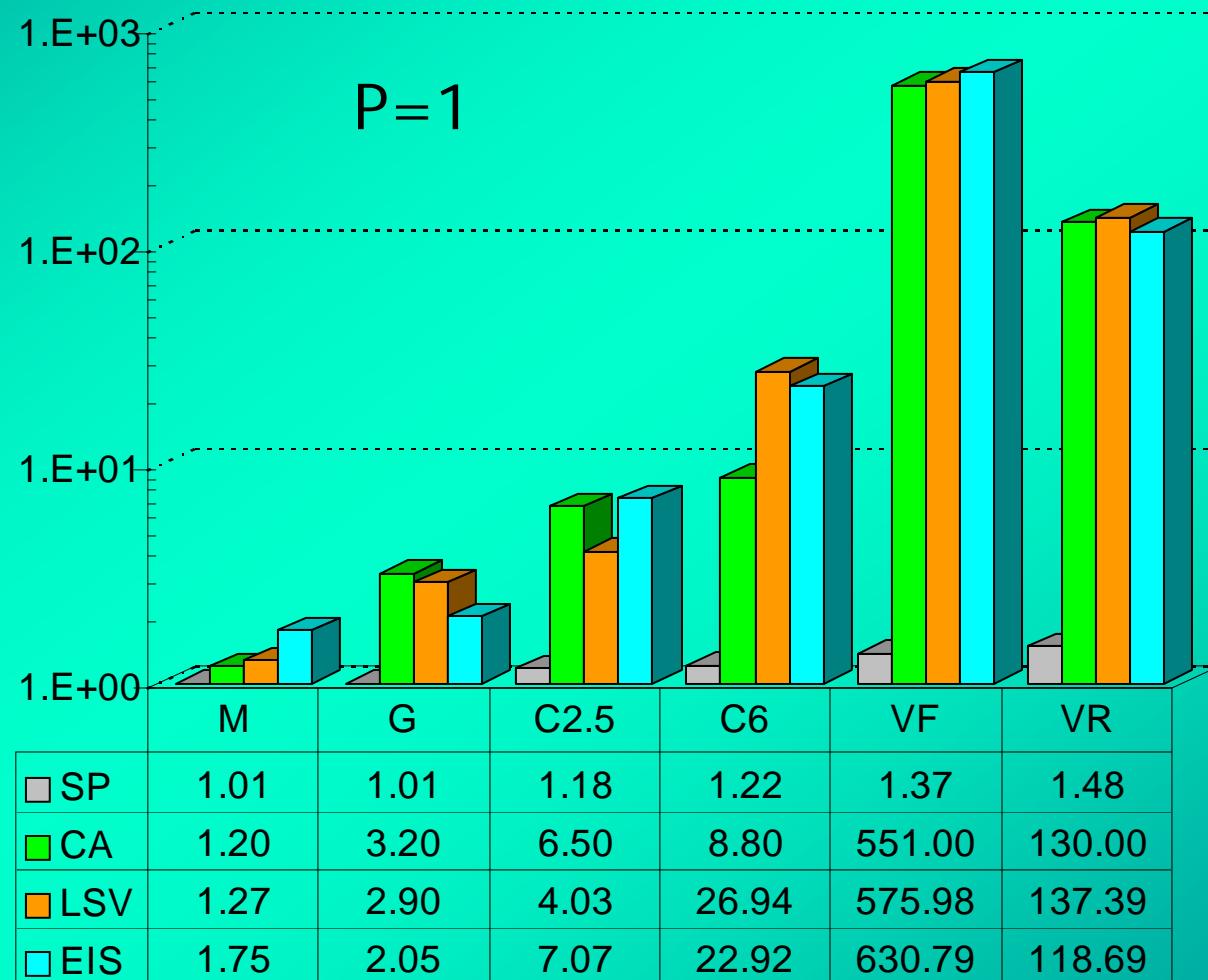
modification	I_{corr} [nA]
P	13
M	16
G	37
C2.5	51
C6	341
VF	7294
VR	1740

Chronoamperometry (CA)



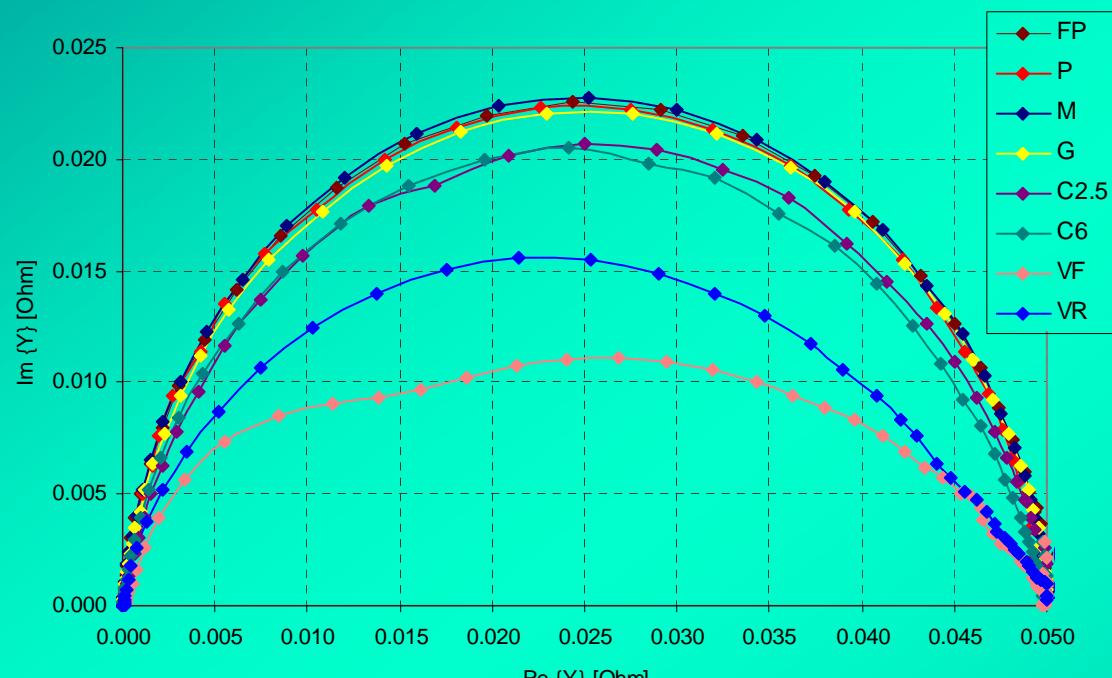
modification	Q [μC]
P	0.86
M	1.03
G	2.75
C2.5	5.59
C6	7.57
VF	474.00
VR	112.00

Increasing of the electroactive area obtained by three electrochemical methods (EIS, LSV, CA) in comparison to that obtained by surface profiling



Determination of the fractal dimension D_F by EIS

Admittance plot for all modifications



FP – fine polished, $\text{Ra}=10 \text{ nm}$

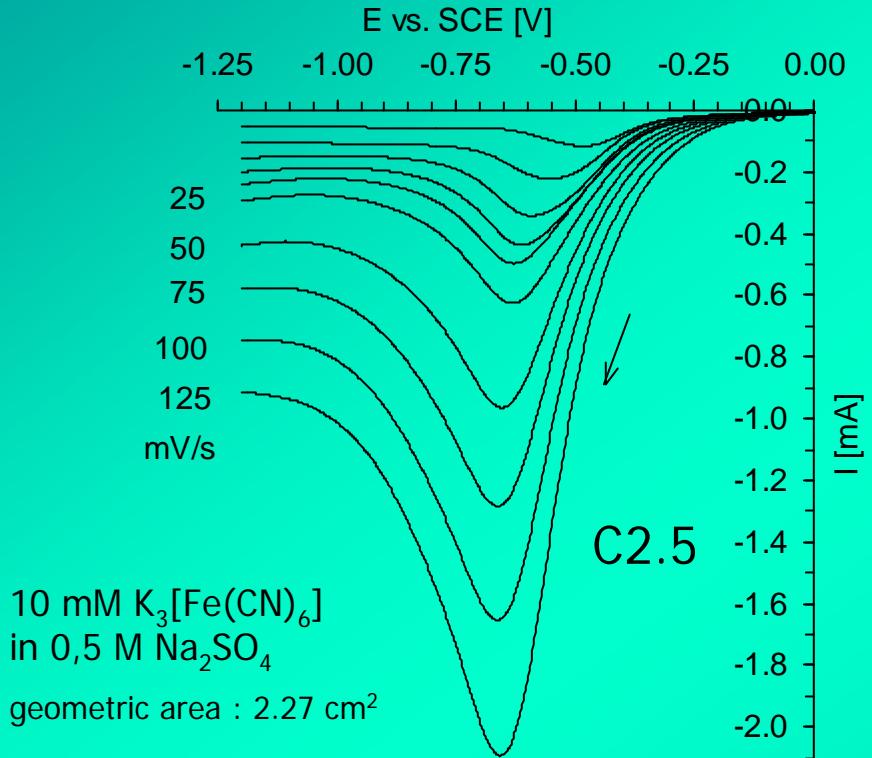
Mr. Velten, University of Saarland, Germany

$$n = \frac{1}{D_F - 1} \quad [1]$$

[1] T. PAJKOSSY, L. NYIKOS
 "Impedance of fractal blocking electrodes"
 J. Electrochem. Soc. 133 (10) (1986) 2061

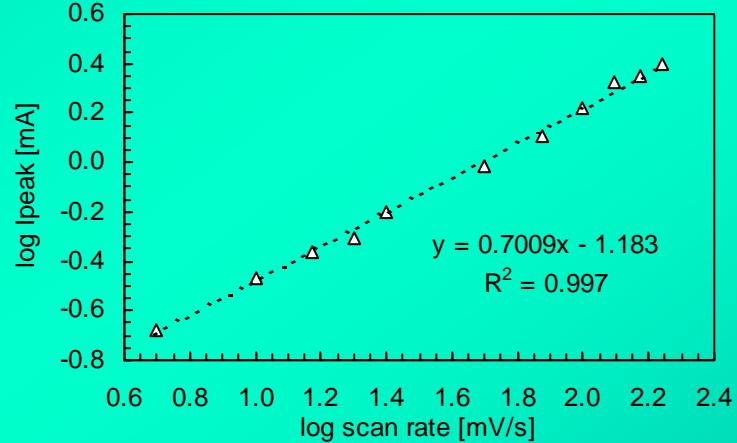
Determination of the fractal dimension D_F by LSV measurement

(reduction of ferricyanide)



$$a = \frac{D_F - 1}{2} \quad [2]$$

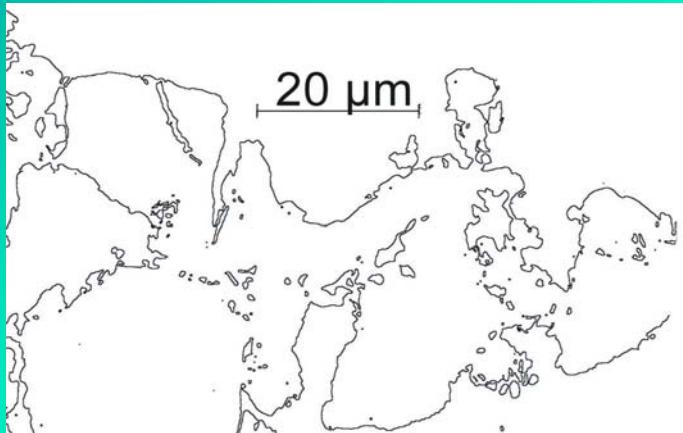
[2] T. PAJKOSSY, L. NYIKOS
"Diffusion to fractal surfaces -
III. Linear sweep and cyclic voltammograms"
Electrochim. Acta 34 (2) (1989) 181-186



modification	slope a
FP	0.598
P	0.642
M	0.745
G	0.574
C2.5	0.701
C6	0.669
VF	0.889
VR	0.769

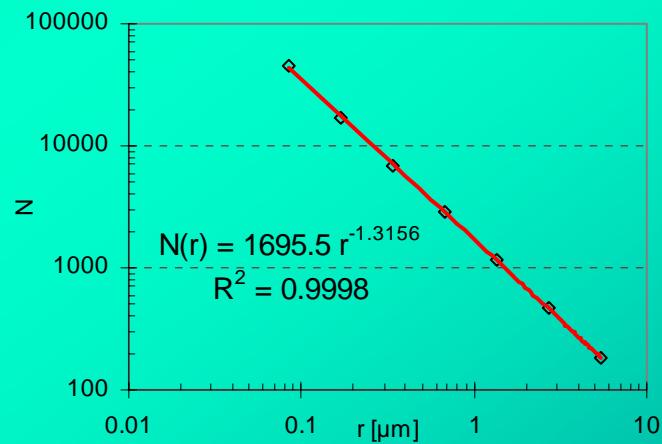
Determination of the fractal dimension D_F by Digital Image Processing (DIP)

Border line obtained from the SEM-picture
of the cross section of a VF-sample



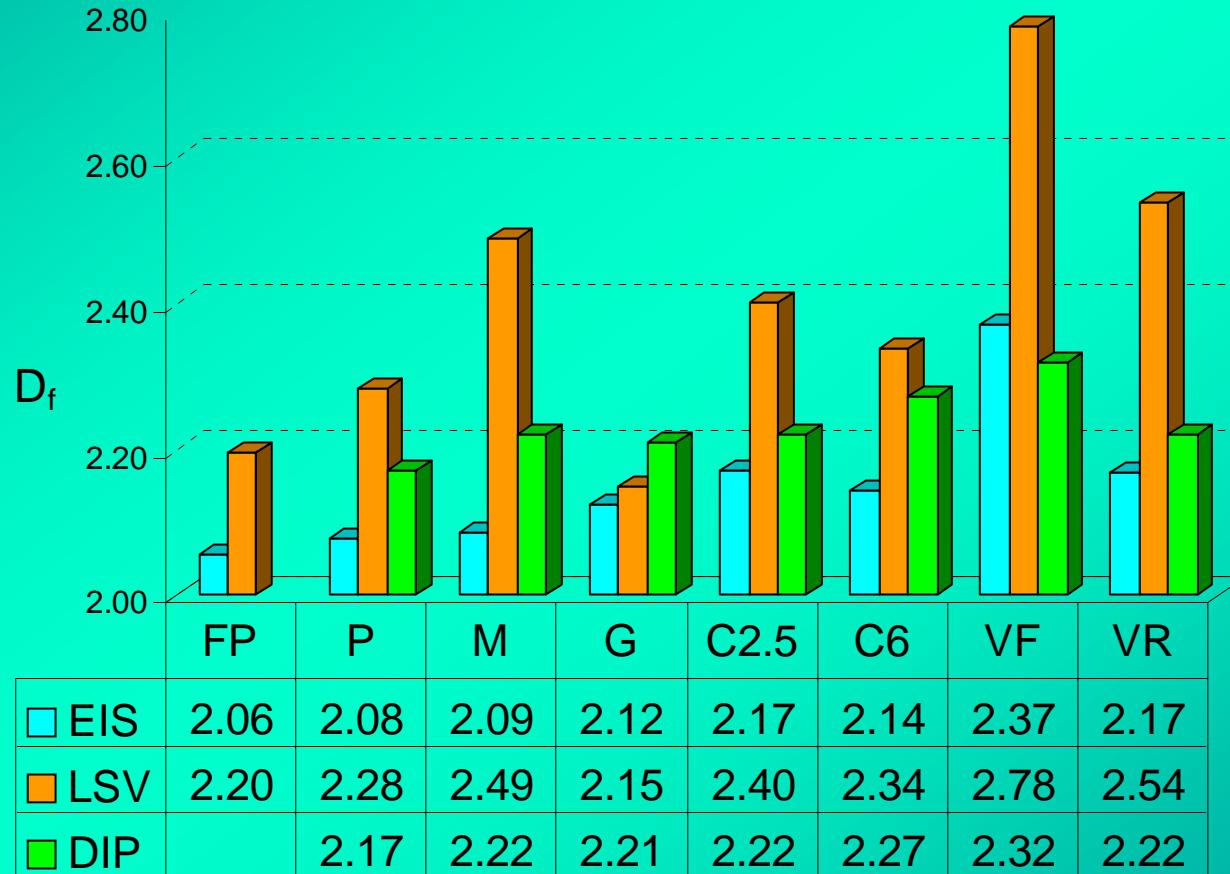
box counting algorithm
(Corel, UTHSCSA Image tool)
 N - Number of boxes
 r - box size
plot $\log(N) = f(\log(r))$
→ slope of the regression line
is the fractal dimension $D_{F,B}$:

Results of the box counting procedure for VF



$$N(r) = \text{const.} \cdot r^{-D_{F,B}}$$

Comparison of the fractal dimension D_F obtained by three different methods



Cell biological examinations

Expression of fibronectin in MG-63 osteoblastic cells
on differently structured titanium surfaces (Western Blot)

M G VR C6 P



R _a [μm]	0.53	1.22	20.99	6.07	0.07
D _F (LSV)	2.49	2.15	2.54	2.34	2.28